This article was downloaded by:

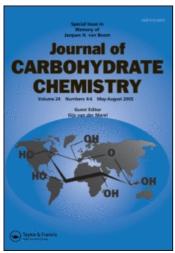
On: 23 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Journal of Carbohydrate Chemistry

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713617200

Communication: A Convenient Synthesis of Per-*O*-Methylated 6-*O*-Monosubstituted ß-Cyclodextrins

Niculina Lupescu; Catherine K. Y. Ho; Guochen Jia; Jiri J. Krepinsky

To cite this Article Lupescu, Niculina , Ho, Catherine K. Y. , Jia, Guochen and Krepinsky, Jiri J.(1999) 'Communication: A Convenient Synthesis of Per-O-Methylated 6-O-Monosubstituted β-Cyclodextrins', Journal of Carbohydrate Chemistry, 18: 1, 99 - 104

To link to this Article: DOI: 10.1080/07328309908543982 URL: http://dx.doi.org/10.1080/07328309908543982

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

COMMUNICATION

A CONVENIENT SYNTHESIS OF PER-O-METHYLATED 6-O-

MONOSUBSTITUTED B-CYCLODEXTRINS

Niculina Lupescu, ¹ Catherine K. Y. Ho, ¹ Guochen Jia, ² and Jiri J. Krepinsky ^{1,2}

¹Department of Medical Genetics and Microbiology, and Protein Engineering Network of Centers of Excellence, University of Toronto, Toronto, Ontario, Canada M5S 1A8;
²Department of Chemistry, Hong Kong University of Science and Technology, Clear Water Bay, Kowloon, Hong Kong.

Received August 3, 1998 - Final Form October 16, 1998

Cyclodextrins are cyclic oligosaccharides that function as chiral host molecules exhibiting enantioselectivity in reactions with racemic guests¹ and thus they represent a significant moiety of artificial enzymes.² The most easily obtainable and relatively inexpensive are β-cyclodextrins (β-CD), cyclic heptaglucosides in which glucopyranosyl residues are linked by an α(1-4) glycosidic bonds. β-CD is surprisingly poorly soluble in water (1.8 g in 100 mL)³ but the solubility of its derivatives is often significantly better. For instance, β-CD with all secondary hydroxyls methylated, forms 50% aqueous solutions at room temperature,⁴ and even completely methylated β-CD is ten times more soluble in water (17 g in 100 mL)⁵ than β-CD itself. Methylation also alters the binding of the substrates to β-CD often increasing its specificity and strength,⁶ and renders the hydroxyl groups unreactive. Therefore, it is of interest to prepare derivatives of β-CD in which the enzyme-mimicking group is bound to one hydroxyl (possibly through a tether) while the rest of the β-CD moiety is permethylated.

100 LUPESCU ET AL.

To synthesize such 6-monosubstituted β-CDs, mono 6-O-tosylates have been used repeatedly⁷ as leaving groups for substitutions with nucleophilic reagents. However, displacement of tosylate with useful nucleophiles usually requires elevated temperatures and long reaction times, yet giving relatively low yields. For instance, it was shown recently that with amines starting from the 6-tosylate, the 6-amino derivatives are obtained in 30-50% yields after 4-8 hours at 70 °C. To Consequently, we decided to explore the use of triflate group, one of the most reactive leaving groups for nucleophilic substitution, to try to improve preparation of the target compounds. In fact, using the triflate of methylated β-CD, substitution with amines occurs almost instantaneously at room temperature giving isolated yields over 75%. Therefore, even less reactive nucleophiles than previously used can be employed.

Mono 6-O-triflate \(\text{B-CD}\) is prepared as depicted in Scheme 1 via a monosilyl derivative. It is well known that alkylation of cyclodextrins with sufficiently bulky agents would preferentially occur at primary hydroxyl groups. In this way mainly single trityl9 or t-butyldimethylsilyl\(^{10}\) derivatives can be obtained. Further methylation of all remaining 20 hydroxyl groups followed by the removal of previously introduced bulky groups yields the desired intermediate. Since the silylation with t-butyldimethylsilyl chloride is accompanied by a number of side products involving multiple silylation, we first tried to suppress the formation of such side products. Expecting that more sterically demanding silylating agents would show higher regiospecificity we examined thexyldimethylsilyl chloride, phenyldimethylsilyl chloride and t-butyldiphenylsilyl chloride in addition to t-butyldimethylsilyl chloride.

Thexyldimethylsilyl chloride and *t*-butyldimethylsilyl chloride gave products of comparable complexity whereas phenyldimethylsilyl chloride gave no reaction. Silylation with *t*-butyldiphenylsilyl chloride provided a cleaner desired product. Thus, silylation and exhaustive methylation of β -CD gave 2, which was converted to 3 upon treatment with NH₄F. Triflate 4 was prepared by treatment of 3 with triflic anhydride and pyridine in dichloromethane in almost quantitative yield. H NMR indicated only the presence of 3 (7-10%) and 4 (90-93%) in the reaction product. One of the most characteristic features of the NMR spectrum of 4 is the downfield shift of H-6 protons at δ 4.92 and δ 4.75 respectively, due to the electron withdrawing effect of the triflate group. ESMS was consistent with the proposed structure for 4 with m/z 1565 [M+NH₄]⁺ and 1570 [M+Na]⁺. The FTIR (nujol) of 4 shows a broad band at 1207 cm⁻¹, characteristic for an S=O stretching bond. The crude crystalline 4 (stable under argon and at -20 °C for several months) was further treated with 2-(2-aminoethoxy)ethanol and ethylenediamine giving the expected substitution products 5 and 6 in excellent yields. Products 5 and 6 thus obtained were fully characterized by H and 13 C NMR spectroscopy and ESMS.

$$(OH)_{7} \qquad (MeO)_{6} \qquad O-Si-R \qquad (MeO)_{6} \qquad OH$$

$$Ph$$

$$Ph$$

$$Ph_{2}(R)SiCl \qquad NH_{4}F \qquad MeOH$$

$$1 \qquad 2 \qquad 3 \qquad Tf_{2}O, Py \qquad CH_{2}Cl_{2}$$

$$Tf = OSO_{2}CF_{3} \qquad (MeO)_{6} \qquad NH-R_{1} \qquad (MeO)_{6} \qquad OTf$$

$$(MeO)_{7} \qquad (OMe)_{7} \qquad (MeO)_{6} \qquad OTf$$

$$F_{1}-CH_{2}-CH_{2}-O-CH_{2}-CH_{2}-OH$$

$$6. \quad R_{1}=CH_{2}-CH_{2}-OH_{2}$$

SCHEME 1

In conclusion, t-butyldiphenylsilyl chloride was found to be the silylating agent of choice yielding the single 6-silyl derivative of β -CD with reasonable regiospecificity. After permethylation, desilylation and chromatography the overall yield of the 6-OH product 3 was 45%. The formation of crystalline triflate 4 and its substitution with bifunctional amines is essentially quantitative.

EXPERIMENTAL

General methods. Thin-Layer Chromatography (TLC) was performed on silica gel plates and visualized by spraying with 50% aq. sulfuric acid and heating at 200 °C. Silica gel (200-400 mesh, Toronto Research Chemicals) was used for flash chromatography. All starting materials were dried overnight in vacuo over KOH or P_2O_5 prior to use, and the solvents were distilled from appropriate drying agents. Solutions were concentrated at 1 mm Hg pressure using a rotary evaporator.

102 LUPESCU ET AL.

¹H and ¹³C NMR spectra were recorded at 500 (125, ¹³C) MHz with a Varian Unity 500 MHz spectrometer at the NMR Spectroscopy Laboratory of the Molecular Medicine Research Centre, University of Toronto. Spectra were obtained either in CDCl₃ containing a trace of TMS (δ=0 ppm, ¹H and ¹³C) as the internal standard or in D₂O (99.98%, Aldrich) containing a trace of acetone (2.225 ppm relative to internal DSS, ¹H and 30.5 ppm ¹³C) as the internal standard using δ(ppm) scale. All mass spectra were recorded with a VG Analytical ZAB-SE mass spectrometer or SCIEX API III mass spectrometer at the Mass Spectrometry Laboratory of the Molecular Medicine Research Centre, University of Toronto. The IR spectra were recorded in a nujol suspension using a Paragon 1000 PC (Perkin Elmer) FTIR spectrometer.

(2,3-Di-O-methyl)hexakis(2,3,6-tri-O-methyl)cyclomaltoheptaose (3). To a solution of dry B-CD (2.27 g, 2 mmol) and imidazole (0.304 g, 3 mmol) in dry DMF (50 mL) was added dropwise, at 70 °C under argon, a solution of tert-butyldiphenylsilyl chloride (1.68 g, 6 mmol) in DMF (2 mL). After stirring for 1 h, at 70 °C, TLC using n-BuOH/EtOH/H₂O:5/4/3¹⁰ indicated that starting β-CD (R_f 0.41) and a faster migrating spot (R, 0.61) were present in the reaction mixture. After cooling to 0 °C, NaH (60% dispersion in mineral oil, 3.2 g, 130 mmol) was added in one portion and the suspension was maintained at 0 °C for 0.5 h and at room temperature for 1 h. Then after cooling to 0 °C, MeI (39 g, 273 mmol) was added slowly over 30 min, and the reaction mixture was allowed to warm to room temperature and stirred overnight. TLC (CHCl₃/MeOH:10/1) showed only spots higher than permethylated B-CD (R, 0.68) indicating that the reaction was complete. The residual NaH was quenched with MeOH (8 mL) at 0 °C, and the resulting mixture was poured into ice water (200 mL) and extracted with CHCl₃ (3 x 75 mL). The chloroform extract was washed successively with Na₂S₂O₃ (3%, 40 mL), water (2 x 30 mL), dried (Na₂SO₄) and concentrated to dryness. The residue was dried in vacuo at 50 °C, dissolved in dry MeOH (100 mL) containing NH₄F (1.1 g), and refluxed for 24 h. After removal of methanol, EtOAc (100 mL) was added, the solids were filtered off using celite, and the filtrate was concentrated. The residue contained two major components R_f 0.68 (permethylated B-CD) and R_f 0.54 (compound 3) (CHCl3/MeOH:10/1). Chromatography on a silica gel column using CHCl3/MeOH:50/1 afforded permethylated B-CD 0.68 g (23%) first, followed by pure 3 (1.27 g; 45%). The NMR and ESMS data of 3 agreed with those published earlier. 10 1H NMR (500 MHz, CDCl₃) δ 5.22 (d, 1H, J = 4.06 Hz, H-1), 5.17 (d, 1H, J = 3.85 Hz, H-1), 5.16 (d, 1H, J = 3.85 Hz, H-1), 5.1 (d, 3H, J = 3.63 Hz, H-1), 5.03 (d, 1H, J = 3.63 Hz, H-1), 3.37-3.89(m, 95H), 3.19 (dd, 7H, J = 3.63 Hz and 9.83 Hz, H-2). ¹³C NMR (125 MHz, CDCl₃) δ 99.02, 98.96, 98.78 (C-1), 82.36, 82.09, 82.02, 81.95, 81.9, 81.8, 81.72, 81.64, 81.59, 81.43, 81.11, 80.67, 80.62, 80.55, 79.96, 79.82, 78.51 (C-2, C-3, C-4), 71.7, 71.62, 71.54,

71.45, 71.33, 71.31, 71.15, 71.0, 70.82 (C-5 plus 6C-6), 61.5 (C-6(OH)), 61.61, 61.5, 61.28, 61.01 (OMe(C-2)), 59.11, 59.02, 58.95, 58.93 (OMe(C-3)), 58.69, 58.62, 58.46, 58.33, 58.24, 58.17 (OMe(C-6)). ESMS *m/z*: 1416 [M+H]⁺, 1433 [M+NH₄]⁺ and 1438 [M+Nal⁺.

(2,3-Di-O-methyl-6-O-trifluoromethanesulfonyl)hexakis(2,3,6-tri-O-methyl) cyclomaltoheptaose (4). To a cold (-15 °C) solution of 3 (300 mg, 0.212 mmol) and pyridine (31.5 μ L) in dry dichloromethane (2 mL), trifluoromethanesulfonic anhydride (90 mg, 0.319 mmol) was added dropwise. The reaction mixture was allowed to warm to room temperature and TLC (CHCl₃/MeOH:10/1) showed that most of the starting material was consumed and a new higher spot (R_f. 0.6) was formed. The solution was cooled again (0 °C), diluted with dichloromethane (10 mL), washed with 5% HCl (1 x 10 mL), aqueous NaHCO₃ (1 x 10 mL) and water (ice cold), dried over Na₂SO₄, filtered and concentrated to dryness to give a crystalline product 4 (325 mg), which was used without purification. Selected NMR data: ¹H NMR (500 MHz, CDCl₃) δ 5.18 (d, 1H, H-1), 5.13 (m, 3H, H-1), 5.105 (d, 1H, H-1), 5.09 (d, 1H, H-1) (all having J = 3.6 Hz), 5.07 (d, 1H, J = 3.85 Hz, H-1), 4.92 (dd, 1 H, J = 1.64 Hz and 10.98 Hz, H-6(OTf)), 4.75 (dd, 1H, J = 6.6 Hz and 10.98 Hz, H-6(OTf), 4.1 (dddd, 1H, J = 1.65 Hz and 6.6 Hz and 8.24 Hz and 10.16 Hz, H-5(OTf), 3.33-3.98 (m, 92H), 3.2 (dd, 7H, J = 3.3 Hz and 9.9 Hz, H-2). ESMS m/z: 1565 [M+NH4]⁺ and 1570 [M+Na]⁺.

{6-Deoxy-2,3-di-O-methyl-6-[2(2-hydroxyethoxy)ethylamino]}hexakis(2,3,6tri-O-methyl)cyclomaltoheptaose (5) and [6-Deoxy-2,3-di-O-methyl-6-(2aminoethyl)amino] hexakis (2,3,6-tri-O-methyl)cyclomaltoheptaose (6). Dry 4 (325 mg), was dissolved in dry DMF (5 mL), cooled to 0 °C, ethylenediamine (120 mg, 2 mmol) was added, and the reaction was allowed to reach room temperature in about 45 min. TLC (CHCl₃/MeOH: 10/1) indicated that 4 was consumed completely and a more polar component (R, 0.2) was formed. The volatiles were removed in vacuo and the residue was chromatographed on silica gel using, CHCl,/MeOH:10/1 to give pure 6, 220 mg (72% overall yield from 3). ¹H NMR (500 MHz, CDCl₃) δ 5.15 (d, 1H, J = 3.66 Hz, H-1), 5.13 (d, 1H, J = 3.42 Hz, H-1), 5.12 (d, 1H, J = 3.54 Hz, H-1), 5.11 (d, 1H, J = 3.54Hz, H-1), 5.1 (d, 1H, J = 3.67 Hz, H-1), 5.08 (d, 1H, J = 3.54 Hz, H-1), 5.02 (d, 1H, J = 3.54 Hz, H-1), 5.02 (d, 1H, J = 3.54 Hz, H-1), 5.02 (d, 1H, J = 3.54 Hz, H-1), 5.03 (d, 1H, J = 3.54 Hz, H-1), 5.04 (d, 1H, J = 3.54 Hz, H-1), 5.05 (d, 1H, J = 3.54 Hz, H-3.66 Hz, H-1), 3.92-3.33 (m, 96H), 3.24 (dd,1H, J = 4.5 Hz and 12.3 Hz, NH (D,O exch)), 3.18 (brdd, 7H, J = 3.42 Hz and 9.9 Hz, H-2), 3.14 (m, 2H, CH₂), 2.8 (m, 2H, CH₂). ¹³C NMR (125 MHz, CDCl₃) δ 99.76, 99.65, 98.94 (C-1), 82.1 (C-2), 81.86, 81.71, 81.44 (C-3), 80.47, 80.28, 80.17 (C-4), 71.36 (C-6), 70.94 (C-5), 61.42, 61.21 (OMe(C-2)), 59.4, 59.00, 58.85 (OMe(C-3)), 58.62, 58.49, 58,33 (OMe(C-6)). ESMS m/z: 1458 $[M+H]^{+}$.

104 LUPESCU ET AL.

In a similar way by using 2-(2-aminoethoxy)ethanol (3 equiv), 5 was obtained in 75% yield. ¹H NMR (CDCl₃) δ 5.15 (d,1H, J = 3.57 Hz, H-1,), 5.12 (m, 4H, H-1), 5.11 (d, 1H, J = 3.57 Hz, H-1), 5.09 (d, 1H, J = 3.57 Hz, H-1), 3.98 (dd, 1H, J = 3.66 Hz and 5.98 Hz), 3.89-3.38 (m, 100H), 3.19 (m, 7H, H-2), 3.15 (t, 2H, CH₂-N). ¹³C NMR (125 MHz, CDCl₃) δ 99.35, 99.16, 98.97, 98.85, 98.74 (C-1), 82.19, 82.12, 82.03, 81.9 (C-2), 81.75, 81.68, 81.61 (C-3), 80.71, 80.46, 80.36 (C-4), 71.00, 70.96, 70.92 (C-5), 71.3 (C-6), 72.38, 61.7 (C-arm), 61.52, 61.48, 61.38, 61.33 (OMe(C-2)), 59.0 (OMe(C-3)), 58.64, 58.58, 58.52, 58.41 (OMe(C-6)). ESMS m/z: 1530 [M+H]⁺ and 1525 [M+Na]⁺

ACKNOWLEDGEMENTS

The spectra were measured in the Molecular Medicine Research Centre partially supported by The Medical Research Council of Canada. The financial support of PENCE and The Hong Kong Research Grant Council is appreciated.

REFERENCES

- C. J. Easton, F. S. Lincoln and D. M.Schliebs, J. Chem. Soc., Chem. Commun., 1167 (1995).
- a) R. Breslow, J. W. Canary, M. Varney, S. T. Waddell and D. Yang, J. Am. Chem. Soc, 112, 5212 (1990); b) Y. Matsumoto and M. Komiyama, Chem. Lett., 469 (1990); c) B. Ekberg, L. Andersson and K. Mosbach, Carbohydr. Res., 192, 111 (1989); d) R. Breslow, A. W. Czarnik, M. Lauer, R. Leppkes, J. Winkler and S. Zimmerman, J. Am. Chem. Soc., 108, 1969 (1986).
- 3. J. Szejtli, in Cyclodextrins and Their Inclusion Complexes, Akadémiai Kiadó, Budapest (1982).
- A. R. Bergeron, Y. Machida, and K. Bloch, J. Biol. Chem. 250, 1223 (1975); R. Breslow, U. Kohn, and B. Siegel, Tetrahedron Lett., 1645 (1976).
- 5. D. French, Adv. Carbohydr. Chem. 12, 189 (1957).
- A. Venema, H. Henderiks and R. V. Geest, J. High Resolut. Chromatogr., 14, 676 (1991); R. P. Villani, S. F. Lincoln and J. H. Coates, J. Chem. Soc., Faraday Trans. 1, 83, 2751 (1987); J. Emert and R. Breslow, J. Am. Chem. Soc., 97, 670 (1975).
- 7. a) G. Wenz, Angew. Chem. Int. Ed. Engl., 33, 803 (1994); b) B. L. May, S. D. Kean, C. J. Easton and S. F. Lincoln, J. Chem. Soc. Perkin Trans. 1, 3157 (1997).
- 8. G. Hodosi and P. Kovac, J. Am. Chem. Soc., 119, 2335 (1997); H. H. Baer and F. Santoyo-Gonzalez, Carbohydr. Res., 280, 315 (1996); L. A. Reed and L. Goodman, Carbohydr. Res., 94, 91 (1981); A. Streitwieser, Jr., C. L. Wilkins and E. Kiehlmann, J. Am. Chem. Soc., 90, 1598 (1968).
- 9. M. Tanaka, Y. Kawaguchi, T. Niinae and T. Shono, J. Chromatogr., 314, 193 (1984).
- 10. Z. Chen, J. S. Bradshaw and M. L. Lee, *Tetrahedron Lett.*, 37, 6831 (1996).
- 11. W. Zhang and M. J. Robins, *Tetrahedron Lett.*, 33, 1177 (1992).